

RECIPES FOR CONFOCAL/ANTIBODY LABELING OF SECTIONS

10X Phosphate Buffered Saline (PBS)

| Chemicals | 250mls | 1Liter | 2Liter |
|----------------------------------|--------|--------|---------|
| NaCl | 21.92g | 87.68g | 175.30g |
| NaH ₂ PO ₄ | 0.67g | 2.68g | 5.3g |
| Na ₂ HPO ₄ | 2.88g | 11.51g | 23.0g |
| NaN ₃ (optional) | 1.25g | 5.0g | 10.0g |

Note: To use the 10X PBS or 10X PBS/NaN₃, stir well before measuring out, dilute with ddH₂O to the desired dilution volume (i.e. 1:10) and adjust pH to 7.4

PBS/Bovine Serum Albumin (BSA)/Triton/Azide ("PBTA")

(dilute antibodies in this solution, and use for rinsing sections)

For 1 Liter, dilute the above 10X solution (with NaN₃) so that 100mls of the 10X solution goes into 900mls ddH₂O, giving the PBS/Azide.

Bovine Serum Albumin (BSA): Add 5g of BSA (Fraction V)

Triton (0.1%): From stock 20% solution, add 5mls to the 1L solution.

Be sure to pH to 7.4

Agarose

(for embedding tissue to vibratome)

Use low gelling agarose (e.g. Sigma's Agarose Type XI low gelling temp, Catalogue # A-3038) in PBS to make a 5% solution with azide. Start with 100 mls PBS, pH 7.3, add 5 g Agarose and 100 ul Na Azide. Heat until dissolved; store in 4°C.

N-Propyl Gallate in Glycerol

(mounting medium for confocal sections)

For 100 mls:

Add 5g N-propyl gallate to 100 mls of glycerol.

Stir overnight with small stir bar.

