

## *Cytochalasin D Disrupts Outer Segment Disc Morphogenesis In Situ in Rabbit Retina*

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Exposure of rabbit retina to cytochalasin D (CD) via a single intraocular injection results in basal rod outer segment (ROS) and cone OS (COS) discs with abnormally large diameters; the overgrown OS membranes extend along either the cell outer or inner segment. Twenty-four hours after the injection, basal ROS and COS discs appear to have recovered their normal diameter, indicating the reversibility of this drug's effect. These data support both the evagination hypothesis for disc morphogenesis and the hypothesis that f-actin's role at the ROS base is to regulate the initiation of membrane evagination and disc diameter. *Invest Ophthalmol Vis Sci* 30:339-342, 1989

The presence of actin at the base of the rod outer segment has been established in a variety of vertebrate species, using antibodies to actin<sup>1</sup>.

*Materials and Methods.* Rabbits were pigmented adults with apparently normal eyes when examined with an indirect ophthalmoscope. They were kept under a 12:12 light/dark regime and the eyes were injected at 3 hr after light onset. One eye of each animal was the experimental eye; the other served as a control. Experimental eyes were injected with 100  $\mu$ l of a stock mixture consisting of 5  $\mu$ g CD (Calbio-chem, San Diego, CA), dissolved to 5 mg/ml in DMSO, plus 995  $\mu$ l of sterile balanced salt solution (BSS; Alcon, Fort Worth, TX). This stock could be stored frozen, but was resterilized by filtration prior to use. Final concentrations of the injected fluids were thus 25  $\mu$ g/ml CD and 0.5% DMSO in sterile BSS. Final concentration in the extracellular space of

brate species, using antibodies to actin<sup>1</sup>.

and the  $\mu$ l

the retina was unknown, but considerably less than

actin-specific probe, fluorescent phalloidin,<sup>3</sup> leading to the suggestion<sup>2</sup> that this contractile protein might perform some major role in disc morphogenesis. New disc membrane is added to the ROS base by evagination of the ciliary membrane.<sup>4</sup> Such membrane movements recall lamellipodial extensions ("ruffles") of motile cells, including growing neurons, which can be attributed to f-actin<sup>5</sup> and may involve a treadmill-ing mechanism.<sup>6</sup>

F-actin's role in membrane movements was established in part with studies employing a family of drugs, the cytochalasins. These compounds bind to the barbed ends of actin microfilaments, inhibiting both association and dissociation of g-actin sub-units.<sup>7,8</sup> The most potent cytochalasin is cytochalasin D (CD), which has the added advantage of not affecting the cell glucose transporter, as do some of the other cytochalasins.<sup>9</sup> While there is still uncertainty as to CD's actual in vivo effects,<sup>10</sup> it is generally accepted that CD inhibits local rearrangements of f-actin that rely on actin polymerization, particularly in cells undergoing shape changes.<sup>11,12</sup>

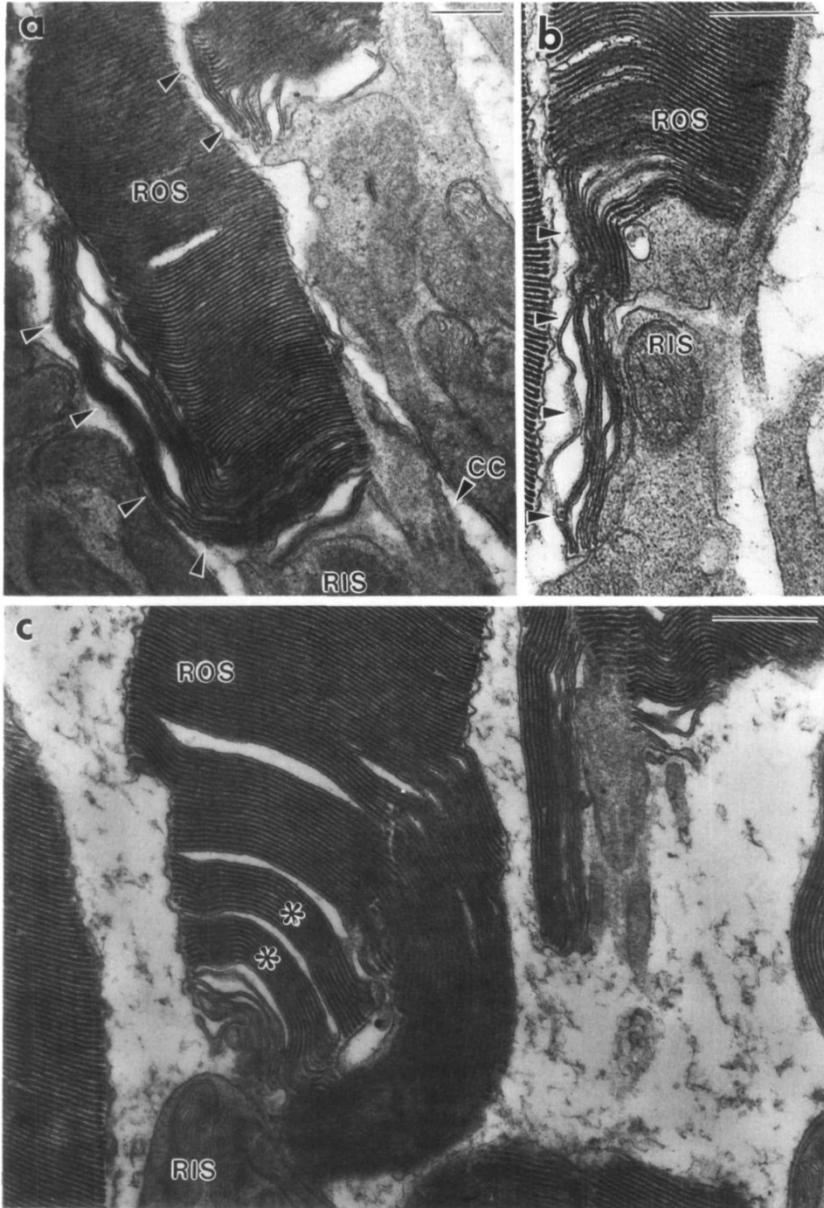
Recently, CD was shown to deregulate ROS disc morphogenesis in *Xenopus* eyecups maintained in vitro.<sup>13</sup> This deregulation was manifested in basal OS discs of abnormally large diameter. We report here that CD administered to the rabbit retina by intraocular injection disrupts OS disc morphogenesis in an identical fashion; moreover, disc diameter appears to recover approximately 24 hr after the drug is injected.

that of the injections themselves.

Rabbits were deeply anaesthetized with an intramuscular injection of Rompun and Ketamine prior to intraocular injections. With the anaesthetized rabbit lying on its side, the sclera of the uppermost eye was gently grasped with a forceps to steady it. A sterile 30-gauge needle was used to penetrate the cornea at its periphery, with care taken to avoid the iris, and then withdrawn. Gentle pressure was applied to the sclera near the corneal incision, draining off some intraocular fluid in order to compensate for the intraocular pressure added by the subsequent injection. The site of injection was through the temporolateral sclera; the needle was inserted approximately 3.5 mm into the posterior vitreous cavity, taking care not to hit the lens. Any sensation of back-pressure during the slow injection was compensated for by another release of fluid from the corneal incision. Following injection, both eyes were given drops of Lidocaine at approximately hourly intervals for 4 hr as the animal recovered from anaesthesia. None of the animals showed any signs of discomfort, nor did any corneal, scleral, or intraocular pathology develop following the procedures. Rompun and Ketamine anaesthesia was repeated prior to euthanasia, which was

accomplished with Nembutal administered by intracardiac puncture. All procedures were performed in accordance with the ARVO Resolution on the Use of Animals in Research.

One animal was killed at each time point (8 or 24



**Fig. 1.** Electron micrographs of rabbit rod photoreceptors, showing the abnormally large diameter (and resulting abnormal displacement) of basal ROS membrane discs after CD injection. ROS = rod outer segment; RIS = rod inner segment; CC = connecting cilium. Scale bars = 5  $\mu$ m. (a) From a retina fixed 8 hr after CD injection. Overgrown basal ROS discs (arrows) extend upward (sclerad) along the ROS plasma membrane. At upper right, a second ROS with overgrown basal discs is seen, but with overgrown discs extending downward (vitread) instead. (b) From a retina fixed 8 hr after CD injection. Overgrown ROS discs extend downward (vitread) along the RIS. (c) From a retina fixed 24 hr after CD injection. Overgrown ROS discs extend downward in two cells. In the cell at left, however, the most basal discs appear to be of a more normal diameter (\*), suggesting that after 24 hr the drug's effect on regulation of disc assembly has been reversed.

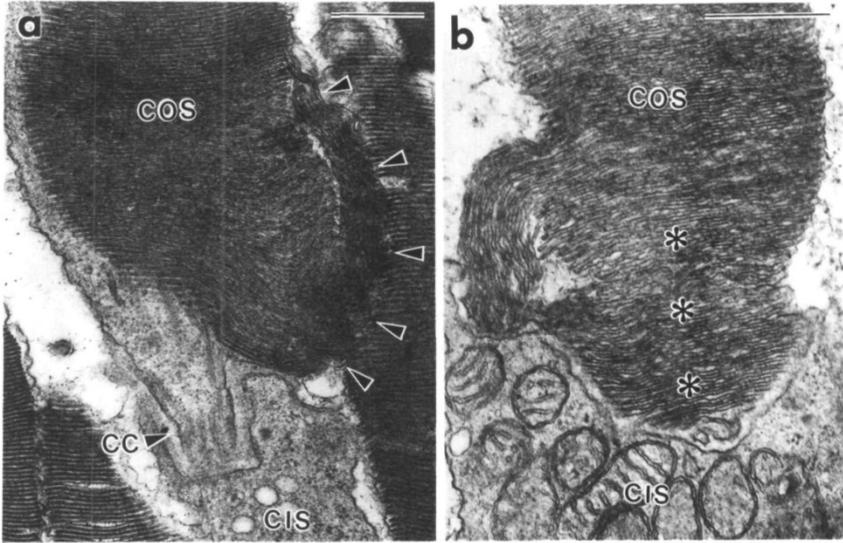


Fig. 2. Electron micrographs of rabbit cone photoreceptors, showing the abnormally large diameter (and resulting abnormal displacement) of basal COS membrane discs after CD injection. COS = cone outer segment; CIS = cone inner segment; CC = connecting cilium. Scale bars = 5  $\mu$ m. (a) From a retina fixed 8 hr after CD injection. Overgrown basal COS discs (arrows) extend upward (sclerad) along the COS. (b) From a retina fixed 24 hr after CD injection. Overgrown COS discs extend downward (vitread) to the CIS. As in the ROSs at 24 hr after injection, the most basal discs appear to have a more normal diameter(\*), suggesting that the drug's effect on regulation of disc assembly has been reversed.

hr post-injection) and the eyes processed for conventional transmission electron microscopy. Eyecups were fixed overnight in 1% paraformaldehyde + 1% glutaraldehyde in phosphate buffer (pH 7.2). After rinsing in buffer, the eyecups were cut into quadrants and postfixed for 1.5 hr with 2% osmium tetroxide in phosphate buffer. Following ethanol dehydration and infiltration with propylene oxide, the quadrants were embedded in Araldite and thin-sectioned on an ultramicrotome (LKB, Bromma, Sweden). Gold or silver sections were collected onto copper mesh grids, stained with uranyl acetate and lead citrate, and examined with a Philips 300 electron microscope (Eindhoven, The Netherlands).

**Results.** In retinas fixed 8 hr after CD injection, most photoreceptor OSs had basal discs with abnormally large diameters, so that the nascent discs appeared to "spill over" the side of the cell. In both rods and cones from CD-treated retinas, overgrown basal OS membranes were observed to extend sclerad along the OS (Figs. 1a, 2a) or vitread along the IS (Figs. 1b, c, 2b). In either case, the overgrown membranes maintained contact with the cell from which they grew. This is in marked contrast to the normal situation in which new discs achieve the diameter of their older, mature counterparts and, following closure,

are displaced further up (sclerad) the ROS. Overgrown discs were not observed in retinas from the control (opposite) eye of any animal.

In retinas fixed 24 hr after CD injection, overgrown discs were still observed in many ROSs and COSs; but the most basal discs had an appearance suggestive of recovery of their normal diameter (Figs. 1c, 2b). The overgrown OS membranes remained apposed to inner or outer segment.

**Discussion.** Williams et al.<sup>13</sup> have previously shown that isolated photoreceptors from *Xenopus* eyecups maintained in CO-supplemented culture medium have f-actin domains that collapse into focal accumulations in the cell body, a typical effect of this drug.<sup>12</sup> The loss of the f-actin domain at the photoreceptor OS base clearly established the sensitivity of that domain to CD. The same study revealed one consequence of that sensitivity, namely that *Xenopus* ROS and COS disc morphogenesis in the presence of CD was abnormal: the basal discs were overgrown, having a larger-than-normal diameter. Concurrent radiolabelling showed that the abnormal discs were assembled during drug exposure, rather than deriving from preexisting discs which were simply altered structurally by the drug. That report<sup>13</sup> thus constitutes the first experimental evidence that actin local-

ized at the OS base is important for disc morphogenesis, and further, provides experimental evidence for the evagination hypothesis of disc morphogenesis proposed by Steinberg and his colleagues.<sup>4</sup>

The present study provides additional evidence for

the evagination hypothesis<sup>4</sup> from an in situ application of the drug and extends the observations on CD's disruptive effect on disc morphogenesis to a mammalian species. Furthermore, the morphology of both ROSs and COSs from rabbit retina fixed 24 hr after

CD injection suggests a recovery of normal disc diameter, i.e., a reversal of the drug's effect on the f-actin domain at the OS base, probably due to washout from the vitreous of what was essentially a pulse of CD (cf. ref. 14). In contrast, the report by Williams et al.<sup>13</sup> was based on continuous delivery of the drug in vitro and correspondingly they did not record any evidence for recovery. We therefore conclude that it is likely that regulation of OS disc morphogenesis by f-actin occurs similarly in both mammals and amphibians and is not an artifact of the in vitro system.

**Key words:** actin, cytochalasin D, disc morphogenesis, photoreceptors, retina

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## References

1. Chaitin MH and Bok D: Immunoferritin localization of actin in retinal photoreceptors. *Invest Ophthalmol Vis Sci* 27:1764, 1986.
2. Chaitin MH, Schneider BG, Hall MO, and Papermaster DS: Actin in the photoreceptor connecting cilium: Immunocytochemical localization to the site of outer segment disc formation. *J Cell Biol* 99:239, 1984.
3. Vaughan DK and Fisher SK: The distribution of f-actin in cells isolated from vertebrate retinas. *Exp Eye Res* 44:393, 1987.
4. Steinberg RH, Fisher SK, and Anderson DH: Disc morphogenesis in vertebrate photoreceptors. *J Comp Neurol* 190:501, 1980.
5. Bray D: Growth cones: Do they pull or are they pushed? *Trends Neurosci* 10:431, 1987.
6. Wang Y-L: Exchange of actin subunits at the leading edge of living fibroblasts: Possible role of treadmill. *J Cell Biol* 101:597, 1985.
7. Brown SS and Spudis JA: Mechanism of action of cytochalasin: Evidence that it binds to actin filament ends. *J Cell Biol* 88:487, 1981.
8. MacLean-Fletcher S and Pollard TD: Mechanism of action of cytochalasin on actin. *Cell* 20:329, 1980.
9. Rampal AL, Pinokofshy HB, and Jung CY: Structure of cytochalasins and cytochalasin B binding site in human erythrocyte membranes. *Biochemistry* 19:679, 1980.
10. Cooper JA: Effects of cytochalasin and phalloidin on actin. *J Cell Biol* 105:1473, 1987.
11. Casella JF, Flanagan MD, and Lin S: Cytochalasin D inhibits actin polymerization and induces depolymerization of actin filaments formed during platelet shape change. *Nature (London)* 293:302, 1981.
12. Schliwa M: Action of cytochalasin on cytoskeletal networks. *J Cell Biol* 92:79, 1982.
13. Williams DS, Linberg KA, Vaughan DK, Fariss RN, and Fisher SK: Disruption of microfilament organization and deregulation of disk membrane morphogenesis by cytochalasin D in rod and cone photoreceptors. *J Comp Neurol* 272:161, 1988.
14. O'Connor P and Burnside B: Actin-dependent cell elongation in teleost retinal rods: Requirements for actin filament assembly. *J Cell Biol* 189:517, 1981.